

Investigating Skin and Soft Tissue Infection (SSTI) mitigation using Far UV-C

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The Problem - Antibiotic dependence and resistance in Skin and Soft Tissue Injuries

- Antimicrobial resistant (AMR) infections are **rapidly increasing** and pose a threat to undo a century of medical progress
- Alternative infection control methods** with broad spectrum effectiveness are desperately needed
- Surgical site infection (SSI) represent **20 % of hospital acquired infections** and patients are at **2-11 times higher risk of death**
- Wound infections require the highest uses of antimicrobial treatments, often due to poor infection control and are the **highest risk of death** beyond the immediate injury
- Antibiotic resistant bacteria, such as CRE (Carbapenem resistant Enterobacteriaceae) has become **resistant to nearly all antibiotics** we have today
- AMR infections threaten **increase the risk of mortality**, result in prolonged patient weakness and **elevate healthcare costs**

The Solution - Far UV-C xIP Infection Protection

- Using Far UV-C (200-235 nm) light to inactivate a broad spectrum of wound site pathogens and including antibiotic resistant bacteria
- Permits reduction in wound site pathogens **without contributing to further antibiotic resistance**
- Far-UVC light allows contactless disinfection which does not physically disrupt the wound site and can be used in conjunction with existing treatments for a **multi-layered approach**

Infections Per Year

Sepsis **1.7M**
Surgery **1.2M**
Chronic Conditions **>30M**
Annual Deaths
> 35 000 in the US
Estimated Cost
> 4.6 B/year and growing
By 2050
1-10 Million Additional Deaths Globally

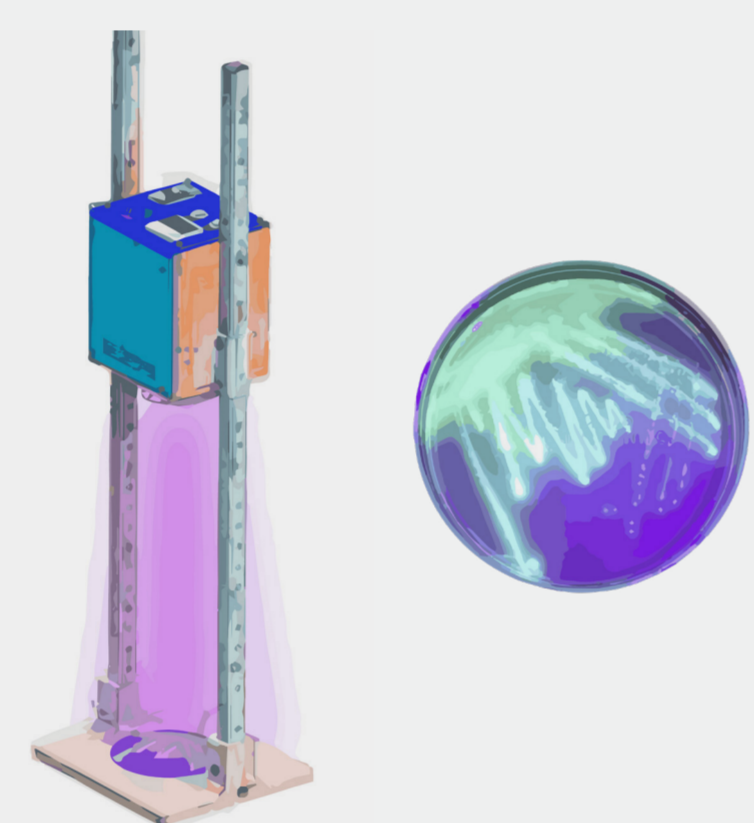
The Investigation - Mitigation in real world cases

- Effectiveness** of different Far UV-C sources including broad spectrum and pulsed sources in reducing MSSA during in-vitro and ex-vivo studies
- Implications of occlusion on **real world wounds** for pathogen reduction
- Penetration and scattering impacts** from expected fluids in wound sites
- Matching of **in-vitro and ex-vivo studies** to actual in-vivo studies carried out with consistent dosing and test methodology

Key Project Studies and Outcomes

In-Vitro (Plate-Based assays)

- Pathogen selection and dose quantification
- UV exposure approach confirmation
- Dose response for each source
- Verifying fluence requirements for pathogens
- Development of chemical dosimetry strategy**
- Validation of fluorescence/luminance dose methods**



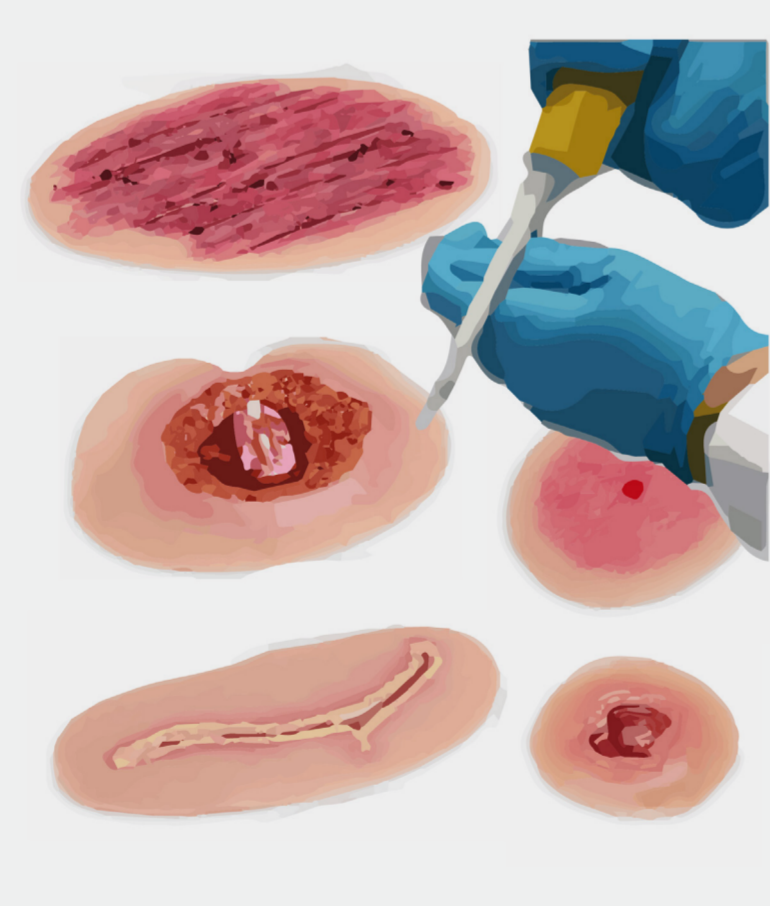
Ex-Vivo (Porcine skin)

- Porcine skin model^x
- Development of wound formation approach
- Development of wound imaging techniques
- Pathogen viability studies performed via CFU
- Validation of fluorescence/luminance dose methods**
- Fluids and effects on fluence, dosimetry and pathogen response**



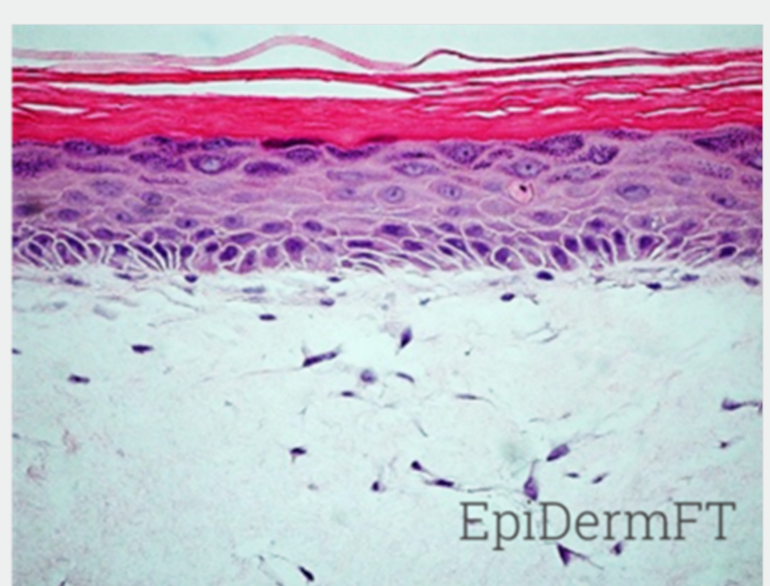
In-Silico Studies (Wound Modeling)

- Establishing theoretical requirements for effective pathogen reduction in wound
- Development of photogrammetry and optical modeling approach
- Fluence modeled in complex wound geometries captured from assays**
- Modeled occlusion and scattering in various fluids**



Ex-Vivo Assays (Human Surrogate)

- Human skin model using EpiDERM FT
- Validation of effective pathogen reduction for reporting via CFU
- Measurement of cellular damage bio-markers (CPD formation)**
- Validation of fluorescence/luminance dose methods**

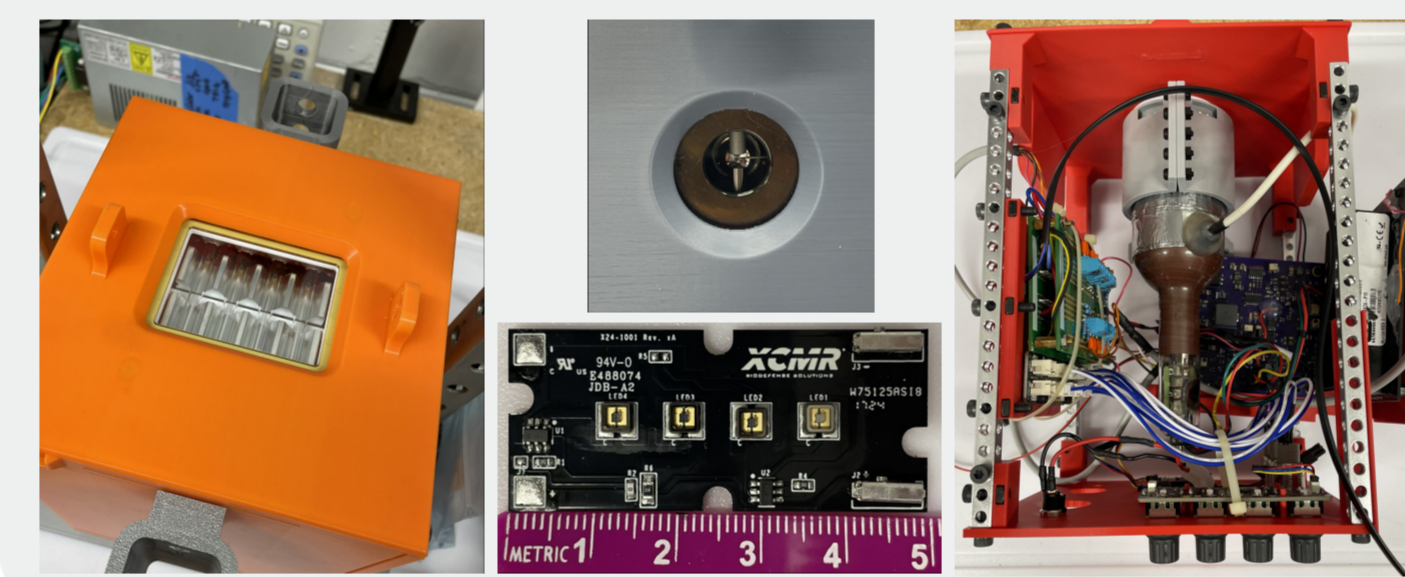
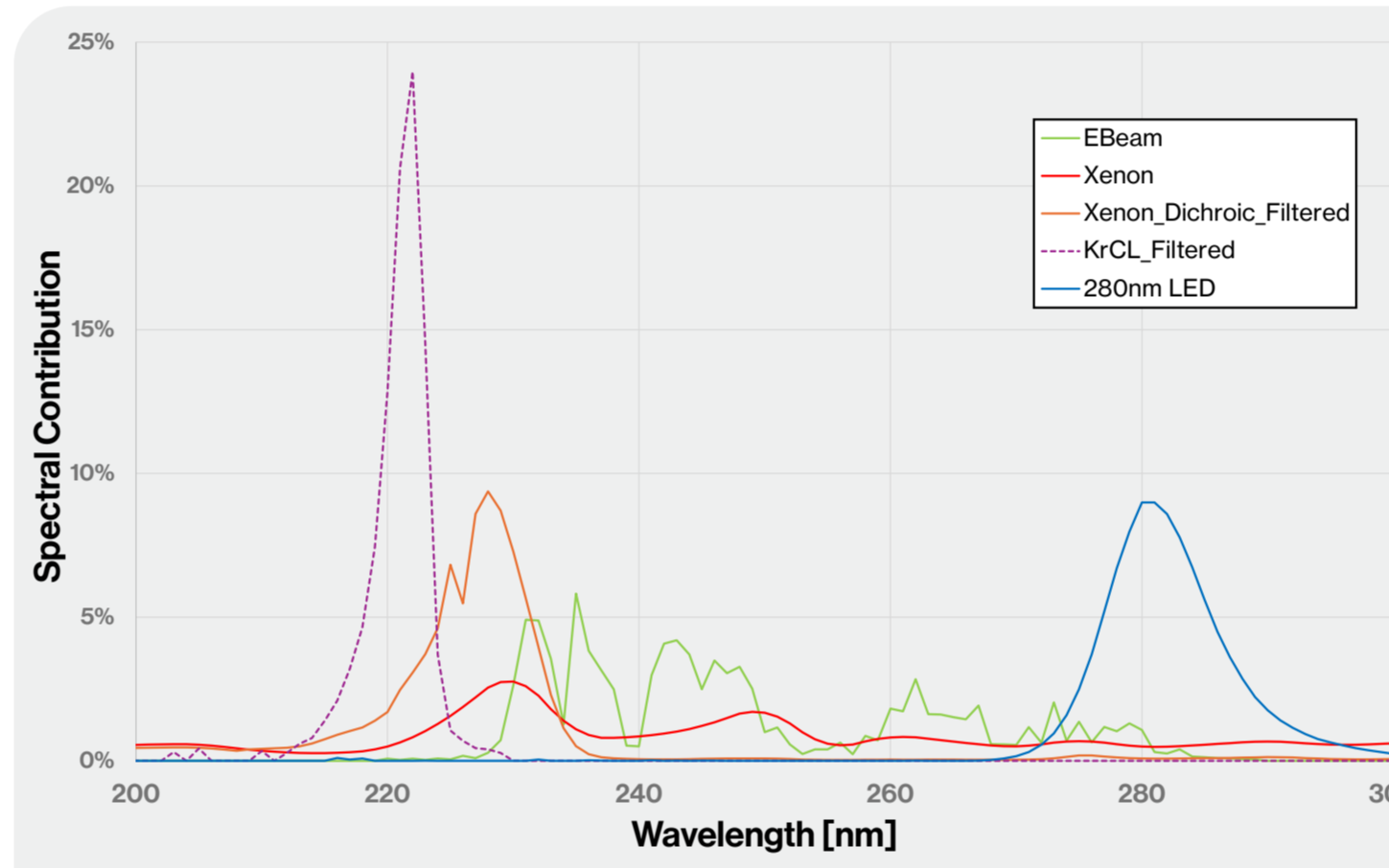


In-Vivo Assays (Mouse Model)

- Nude rodent model with humanized skin (SKH1 Elite)
- Live pathogen testing with MSSA or appropriate surrogate
- Pathogen reduction characterized via CFU*
- Longitudinal studies with fluorescent or bioluminescent cells showing UV dose**



Project Work Completed / In-Progress

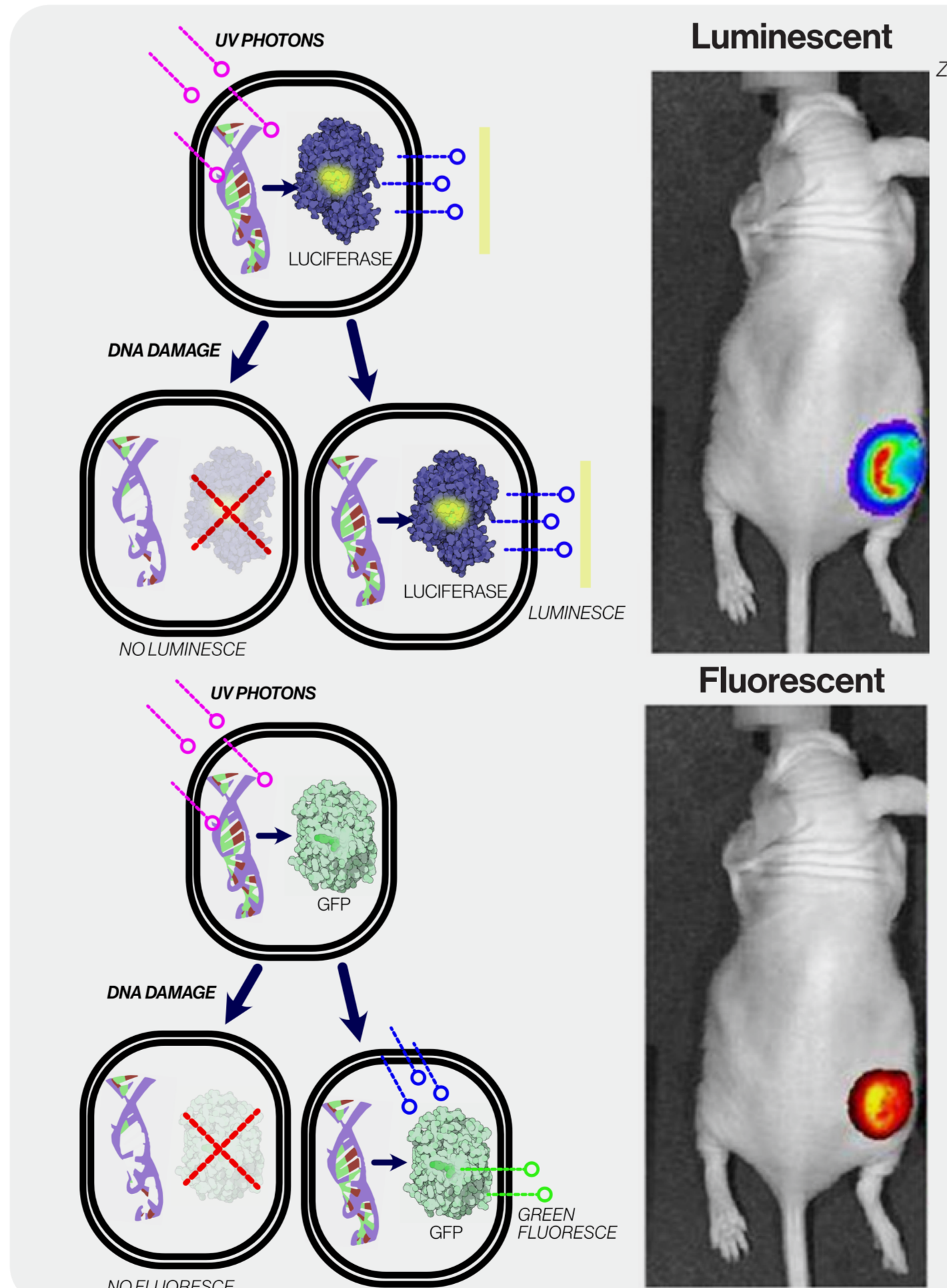


UV-C Sources Studied

- Range of sources used in the study including:
 - KrCl Excimer (222nm peak)
 - Electron Beam Cathodoluminescent (235nm peak)
 - Pulsed Xenon (230nm peak)
 - 280nm LED (280nm peak)
- Includes both pulsed, multi-spectrum sources (EBeam CL and Xenon) and continuous narrow spectrum sources (KrCL and LED)
- Only currently available commercial or high TLR sources used in the study
- Equivalent dose and equivalent ACIGH TLV limit (skin) optical outputs used as limits of study

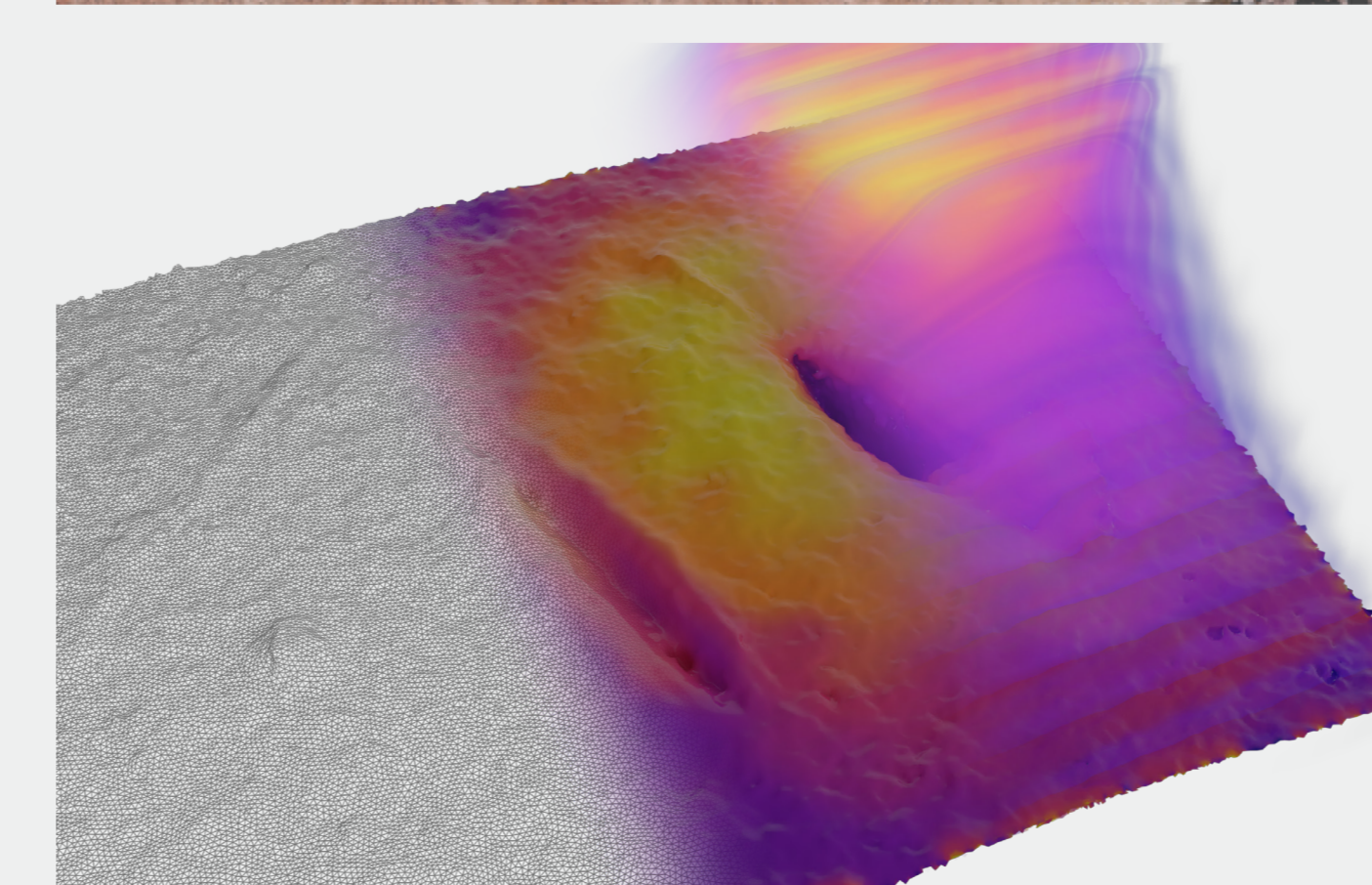
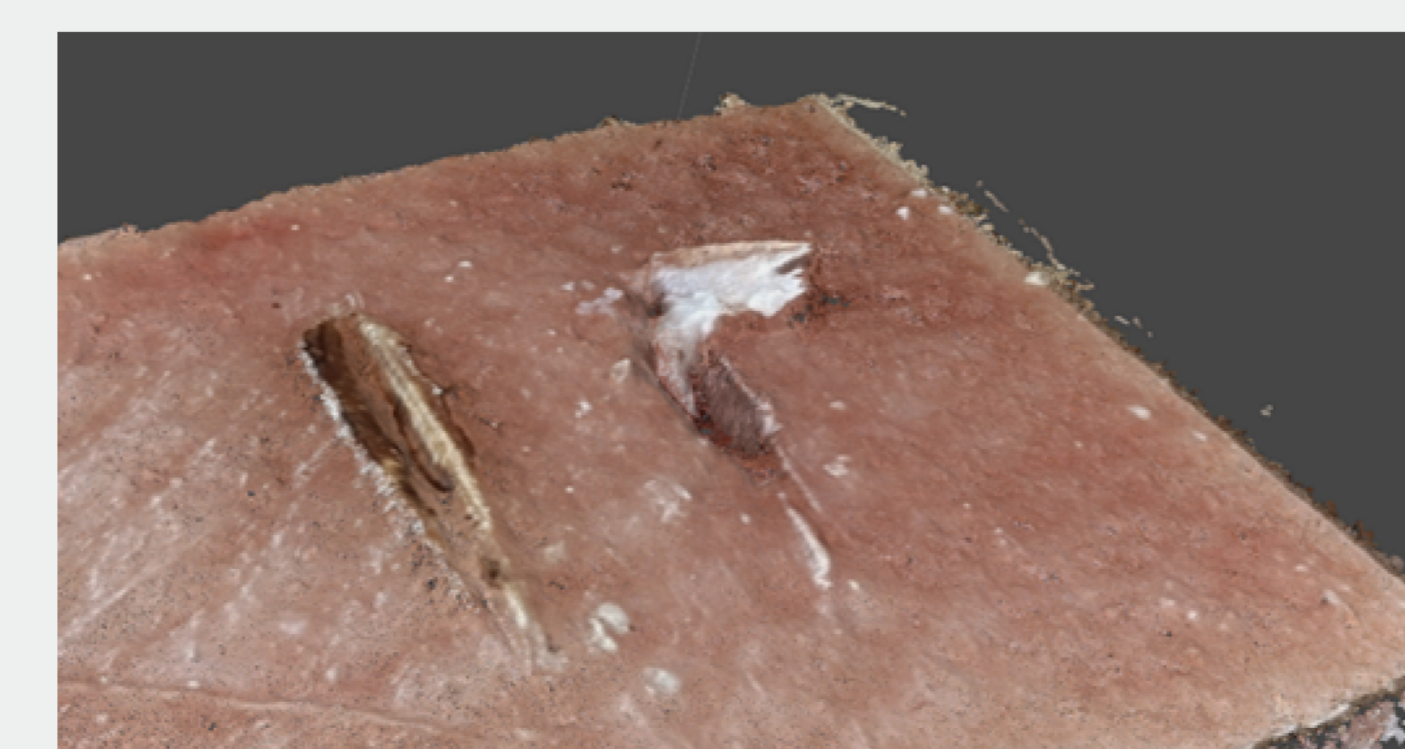
Temporal Dose Estimation

- Studies using engineered MSSA strains which either luminescent or fluorescent reporters
- Viable cells will continue to luminesce/fluoresce giving indication of successful gene expression
- Testing with reporters underway to understand UV-C response:
 - Establishment of best process (luminescent or fluorescent)
 - Demonstration of quenching / decay rate under UV-C exposure and control plates
 - Establishment of dose response versus luminescence/fluorescence of the strains versus CFU
 - Repeatability and ex-vivo suitability/performance studies
- Potential imaging with IVIS as part of in-vivo assays to determine spatial distribution of inactivation



Wound Modeling

- Wounds created to mimic multiple injury types and sizes for ex-vivo (Porcine) studies
- Actual wound geometry results in occlusion, scattering and altered penetration compared to plate assays
- In-silico studies are used to determine best approach including:
 - Source view angle and view factor
 - Traversal/multiple exposure requirements
- 3D geometry for created wounds will be created for simulation using macro photogrammetry
 - Multiple images taken of wound site at different distance and angles
 - Depth estimation and point clouds from images
 - Re-triangulation of point cloud to determine geometry
- Geometry can be used directly for in-silico experiments



* Future Studies
X. M. A. Andersson, L. B. Madsen, A. Schmidtchen, and M. Puthia, "Development of an Experimental Ex Vivo Wound Model to Evaluate Antimicrobial Efficacy of Topical Formulations," Int. J. Mol. Sci., vol. 22, no. 9, p. 5045, Y. T. N. Demidova, F. Gad, T. Zahra, K. P. Francis, and M. R. Hamblin, "Monitoring photodynamic therapy of localized infections by bioluminescence imaging of genetically engineered bacteria," J. Photochem Z. - Vis Optical Imager, THE OLIVE LABORATORY, Accessed: Apr. 12, 2024. [Online]. Available: https://www.OliveLab.org/ivis-optical-imager.html